

REPORT DOCUMENTATION PAGE

Form Approved OMB NO. 0704-0188

Public reporting burden for this collection of information is estimated to average 1 hour per response, including the time for reviewing instructions, searching existing data sources, gathering and maintaining the data needed, and completing and reviewing the collection of information. Send comment regarding this burden estimates or any other aspect of this collection of information, including suggestions for reducing this burden, to Washington Headquarters Services. Directorate for information Operations and Reports, 1215 Jefferson Davis Highway, Suite 1204, Arlington, VA 22202-4302, and to the Office of Management and Budget. Paperwork Reduction Project (0704-0188), Washington, DC 20503.

1. AGENCY USE ONLY (Leave blank)

2. REPORT DATE 1/02-01

3. REPORT TYPE AND DATES COVERED FINL REPORT 15 SA 95- 31 Dec, 98

4. TITLE AND SUBTITLE

Neurobiological Correlates of Sleep Homeostasis

5. FUNDING NUMBERS

6. AUTHOR(S)

Thomas S. Kilduff, Ph. D.

7. PERFORMING ORGANIZATION NAMES(S) AND ADDRESS(ES)

The Board of Trustees of theLeland Stanford Junior Univ. Sponsored Projects Office

857 Serra Street. Room 260 Stanford, CA 94305-4125

9. SPONSORING / MONITORING AGENCY NAME(S) AND ADDRESS(ES)

U.S. Army Research Office P.O. Box 12211 Research Triangle Park, NC 27709-2211 8. PERFORMING ORGANIZATION REPORT NUMBER

10. SPONSORING / MONITORING AGENCY REPORT NUMBER

ARO34616.2-LS

11. SUPPLEMENTARY NOTES

The views, opinions and/or findings contained in this report are those of the author(s) and should not be construed as an official Department of the Army position, policy or decision, unless so designated by other documentation.

12a. DISTRIBUTION / AVAILABILITY STATEMENT

12 b. DISTRIBUTION CODE

Approved for public release; distribution unlimited.

20010222 038

13. ABSTRACT (Maximum 200 words)

This research was intended to examine molecular aspects of sleep homeostasis in the brain. Our experiments were guided by the two-process model of sleep regulation which posits that an increased homeostatic "drive" to sleep occurs during prolonged wakefulness. We found that expression of brain-derived neurotrophic factor (BDNF) mRNA increases in the rat cortex during a 6 hr sleep deprivation period. Increased BDNF mRNA levels likely results in elevated expression of BDNF protein which we hypothesize may protect neurons from the potentially deleterious effects of prolonged sensory stimulation during wakefulness.

In the course of these studies, we cloned a novel gene, hypocretin, that encodes two biologically active neuropeptides expressed within a very restricted area of the posterior hypothalamus. When injected into the lateral venticles of the brain, the hypocretin peptides stimulate food intake, increase wakefulness and decrease deep slow wave sleep and REM sleep. In humans, the sleep disorder narcolepsy has recently been associated with degeneration of the hypocretin neurons. We have therefore proposed a model in which the hypocretins play a central role in arousal state (i.e., sleep/wake) regulation.

We also studied gene expression in the hibernating brain, as another model of arousal state.

14. SUBJECT TERMS

Sleep, sleep deprivation, brain, genes, neuropeptide, hypocretin, orexin, hibernation

15. NUMBER IF PAGES 5 + COVER

16. PRICE CODE

17. SECURITY CLASSIFICATION OR REPORT UNCLASSIFIED 18. SECURITY CLASSIFICATION OF THIS PAGE UNCLASSIFIED 19. SECURITY CLASSIFICATION OF ABSTRACT UNCLASSIFIED

20. LIMITATION OF ABSTRACT

UL

(4) STATEMENT OF THE PROBLEM STUDIED

The purpose of this proposal was to examine the biochemical and molecular aspects of sleep homeostasis in the brain. The experiments were guided by the two-process model of sleep regulation which posits that an increased homeostatic "drive" to sleep occurs during prolonged wakefulness. We hypothesize that (1) a molecular basis exists for the homeostatic regulation of sleep; (2) perturbations of the sleep homeostatic system result in compensatory changes in gene expression in brain that increase the likelihood of subsequent sleep; and (3) sleep after prolonged wakefulness involves a change in macromolecular synthesis that facilitates neuronal recovery or restoration.

In the course of pursuing this hypothesis, we identified a novel hypothalamic gene which encoded two previously undescribed neuropeptides. A large part of our effort during the funding period was devoted to studying the anatomy and physiology of this novel neurotransmitter system. The relevance of this work to sleep has been established by recent studies which indicate that ICV injections of the hypocretins promote wakefulness and decrease deep slow wave sleep and REM sleep and the discovery that, in humans, the sleep disorder narcolepsy is associated with degeneration of the hypocretin neurons.

(5) SUMMARY OF THE MOST IMPORTANT RESULTS

<u>Sleep deprivation.</u> During the course of this research, we found that expression of brain-derived neurotrophic factor (BDNF) mRNA increases in the cortex during a 6 hr sleep deprivation period (Peyron et al., 1998b). Increased BDNF mRNA levels in the cortex likely results in elevated expression of BDNF protein which we hypothesize may protect neurons from the potentially deleterious effects of sensory stimulation during wakefulness.

<u>Isolation of the hypocretin gene</u>. Using directional tag PCR subtractive hybridization (Kilduff et al., 1998), we isolated a novel neuropeptide gene, hypocretin, that encodes biologically active

two peptides, hypocretin-1 and hypocretin-2 in a collaboration with Dr. Greg Sutcliffe at The Scripps Research Institute (de Lecea et al., 1998). *In situ* hybridization revealed that preprohypocretin was expressed exclusively by a bilaterally symmetric structure within the posterior hypothalamus. This hypothalamus–specific mRNA was the precursor of a pair of peptides which share substantial amino acid identities with the gut hormone secretin. Rat preprohypocretin encodes a 130 amino acid putative secretory protein with 4 sites for potential proteolytic maturation. Two of the putative products, which appear to be C–terminally amidated, share 14 identities across 20 amino acids. This region shares a 7/7 match with the gut hormone secretin. Mouse hypocretin (*Hcrt*) differs from rat at 7 amino acids. These differences obliterate 2 of the possible proteolytic products, but preserve the peptides that are related both to each other and to secretin, consistent with an evolutionarily conserved function for the 2 new peptides. The *Hcrt* gene, which in mouse is

located on chromosome 11, is expressed most prominently after postnatal week 3.

In situ hybridization indicates that the neurons expressing preprohypocretin mRNA are located exclusively in the posterior hypothalamus. A polyclonal antiserum was raised in rabbits against the C-terminal 17-residues of preprohypocretin. We undertook an immunohistochemical study to determine the distribution of preprohypocretin-immunoreactive (preprohypocretin-IR neurons and fibers in the brain (Peyron et al., 1998a). In accord with the *in situ* hybridization data, preprohypocretin-IR cell bodies were observed exclusively in the perifornical nucleus and the dorsal and lateral hypothalamic areas. The fibers of these neurons are widespread throughout the posterior hypothalamus and project to multiple targets in other areas, including brainstem and thalamus. Preprohypocretin-IR fibers were located throughout the posterior hypothalamus, and in the preoptic area, the mediodorsal and reuniens nuclei of the thalamus, the dorsal raphe nucleus, the locus coeruleus, the laterodorsal tegmental nucleus, the central gray, the colliculi and the nucleus of the solitary tract. Few labeled fibers were located in cortical regions. These results indicate that preprohypocretin is translated as a peptide in the rat hypothalamus and transported to several brain areas.

At the EM level, preprohypocretin-IR is associated with large granular vesicles at synapses. One of the Hcrt peptides (hcrt-2) was excitatory when applied to cultured, synaptically coupled

hypothalamic neurons, but not hippocampal neurons (de Lecea et al., 1998). These observations suggest that the hypocretins function within the CNS as neurotransmitters. Indeed, another group subsequently cloned the same gene and the cognate receptors for the two peptides which they called the <u>orexins</u> because, when injected into the lateral venticles of the brain, these peptides stimulated food intake(Sakurai et al., 1998). We subsequently determined that one-third of all medial and lateral hypothalamic neurons tested, but not hippocampal neurons, showed a striking nanomolar sensitivity to hypocretin (van den Pol et al., 1998). As studied with calcium digital imaging with fura-2, hypocretin raised cytoplasmic calcium via a mechanism based on G-protein enhancement of calcium influx through plasma membrane channels. The peptide has a potent effect at both presynaptic and postsynaptic receptors. With whole-cell patch-clamp recording, we showed that hypocretin, acting directly at axon terminals, can increase the release of the amino acid transmitters GABA and glutamate. We also showed that Hcrt-2 regulates the synaptic activity of physiologically identified neuroendocrine neurons studied in hypothalamic slices containing the arcuate nucleus, suggesting a function of hypocretin in hormone regulation.

The most dense arborization of hypocretin axons in the brainstem was detected in the locus coeruleus (LC). In electrophysiological studies with slices of rat brain, we found that all LC cells showed excitatory responses to the hypocretin-2 peptide (Horvath et al., 1999). Hypocretin-2 uniformly increased the frequency of action potentials in these cells, even in the presence of tetrodotoxin, indicating that receptors responding to hypocretin were expressed in LC neurons. Two mechanisms for the increased firing rate appeared to be a reduction in the slow component of the afterhyperpolarization (AHP) and a modest depolarization. Our observations suggest a signaling pathway via which signals acting on the lateral hypothalamus may influence the activity of the LC and thereby a variety of CNS functions related to noradrenergic innervation, including vigilance, attention, learning, and memory. Thus, the hypocretin innervation of the LC may serve to focus cognitive processes to complement hypocretin-mediated activation of autonomic centers already described.

The relevance of this work to sleep has been established by recent studies which indicate that ICV injections of the hypocretins promote wakefulness and decrease deep slow wave sleep and REM sleep (Hagan et al., 1999; Piper et al., 2000). Furthermore, two groups have recently shown that, in humans, the sleep disorder narcolepsy is associated with degeneration of the hypocretin/orexin neurons (Peyron et al., 2000; Thannickal et al., 2000). We have recently integrated this information into a general model for the role of the hypocretins in arousal state regulation (Kilduff and Peyron, 2000).

Gene Expression and Mammalian Hibernation. Very little information is available on molecular changes that correlate with hibernation state, and what has been done focused mainly on seasonal changes in peripheral tissues. The purpose of this study (O'Hara et al., 1999) was to characterize changes in gene expression in the brain of a seasonal hibernator, the golden-mantled ground squirrel, Spermophilus lateralis, during the hibernation season. We produced over 4000 reverse transcription-PCR products from euthermic and hibernating brain and compared them using differential display. Twenty-nine of the most promising were examined by Northern analysis. Although some small differences were observed across hibernation states, none of the 29 had significant changes. However, a more direct approach, investigating expression of putative hibernation-responsive genes by Northern analysis, revealed an increase in expression of transcription factors c-fos, junB, and c-Jun, but not junD, commencing during late torpor and peaking during the arousal phase of individual hibernation bouts. In contrast, prostaglandin D2 synthase declined during late torpor and arousal but returned to a high level on return to euthermia. Other genes that have putative roles in mammalian sleep or specific brain functions, including somatostatin, enkephalin, growth-associated protein 43, glutamate acid decarboxylases 65/67, histidine decarboxylase, and a sleep-related transcript SD464 did not change significantly during individual hibernation bouts. We did not observe a decline in total RNA or total mRNA during torpor as others had been previously hypothesized. Therefore, it appears that the dramatic changes in body temperature and other physiological variables that accompany hibernation involve only modest reprogramming of gene expression or steady-state mRNA levels.

We also examined expression of heat shock protein 70 (HSP70) in greater detail (Bitting et al., 1999). RNA transcripts of 2.7 and 2.9 kb hybridizing to an HSP70 cDNA were expressed in both brain and peripheral tissues of pre-hibernation euthermic animals; higher levels of expression were observed during the day than during nighttime samples. A decline in the expression of both transcripts occurred in all tissues examined during hibernation that remained low throughout the hibernation season, including the interbout euthermic periods and regardless of time of day. Quantitative comparisons showed pre-hibernation nighttime HSP70 expression to be as low as that observed during hibernation, despite the drastic increase in metabolic state and nearly 30 degrees C difference in body temperature. In contrast to HSP70, some mRNAs, such as beta-actin and HSP60, remained relatively constant, while others, such as glyceraldehyde 3-phosphate dehydrogenase, increased in specific tissues during the hibernation season. These results indicate that the expression of a highly conserved gene involved in protection from cellular stress, HSP70, can vary with an animal's arousal state.

(6) LIST OF ALL PUBLICATIONS AND TECHNICAL REPORTS:

- Kilduff, T. S., L. de Lecea, H. Usui and J. G. Sutcliffe (1998). Isolation and identification of specific transcripts by subtractive hybridization. Pp. 103-118 in *Molecular Regulation of Conscious States*, R. Lydic, ed. Boca Raton: CRC Press.
- de Lecea*, L., T.S. Kilduff*, C. Peyron, X.-B. Gao, P.E. Foye, P.E. Danielson, C. Fukuhara, E.L.F. Battenberg, V.T. Gautvik, F.S. Bartlett II, W.N. Frankel, A.N. van den Pol, F.E. Bloom, K.M. Gautvik, and Sutcliffe, J.G. (1998). The hypocretins: hypothalamus-specific peptides with neuroexcitatory activity. *Proceedings of the National Academy of Sciences* 95: 322-327. (http://www.pnas.org/cgi/content/abstract/95/1/322)
- van den Pol, A.N., X.-B. Gao, K. Obrietan, T.S. Kilduff and A.B. Belousov (1998). Presynaptic and postsynaptic actions and modulation of neuroendocrine neurons by a new hypothalamic peptide, hypocretin/orexin. *Journal of Neuroscience* 18: 7962-7971. (http://www.jneurosci.org/cgi/content/full/18/19/7962)
- Kilduff, T.S. and C. A. Kushida (1998). Circadian regulation of sleep. Pp. 135-147 in *Sleep Disorders Medicine: Basic Science, Technical Considerations and Clinical Aspects.* S. Chokroverty, ed. Boston: Butterworth Heinemann Publishers.
- Peyron, C., D.K. Tighe, A.N. van den Pol, L. de Lecea, H.C. Heller, J.G. Sutcliffe and T.S. Kilduff (1998). Neurons containing hypocretin (orexin) precursor project to multiple neuronal systems. *Journal of Neuroscience* 18:9996-10015. (http://www.jneurosci.org/cgi/content/full/18/23/9996).
- Kilduff, T.S. and E. Mignot (1999). Molecular and genetic aspects of sleep. Pp. 343-367 in *Neurobiology of Sleep and Circadian Rhythms*, F. W. Turek and P. Zee, eds. New York: Marcel Dekker, Inc.
- O'Hara, B.F., F.L. Watson, H. Srere, H. Kumar, S.W. Wiler, S.K. Welch, L. Bitting, S.K. Welch, H.C. Heller and T.S.Kilduff (1999). Gene expression in brain across the hibernation cycle. *J. Neurosci.* 19:3781-3790. (http://www.jneurosci.org/cgi/content/full/19/10/3781)
- Bitting, L., F. L. Watson, B.F. O'Hara, T. S. Kilduff, and H. Č. Heller (1999). HSP70 expression is increased during the day in a diurnal animal, the golden-mantled ground squirrel *Spermophilus lateralis*. *Molecular and Cellular Biochemistry* 199: 25-34.
- Horvath, T. L., C. Peyron, S. Diano, A. Ivanov, G. Aston-Jones, T. S. Kilduff, A. N. van den Pol (1999). Hypocretin (orexin) and synaptic innervation of the locus coeruleus noradrenergic system. *J. Comp. Neurol.* 415:145-159

(6B) LIST OF ABSTRACTS:

Sutcliffe, J.G., K.M. Gautvik, T.S. Kilduff, T. Horn, P.E. Foye, P.E. Danielson, W.N. Frankel, F.E. Bloom and L. de Lecea (1997). Two novel hypothalamic peptides related to secretin derived from a single neuropeptide precursor. *Society for Neuroscience Abstracts* 23: 2032.

- Peyron, C., D.K. Tighe, B.S. Lee, L. de Lecea, H.C. Heller, J.G. Sutcliffe and T.S. Kilduff (1997). Distribution of immunoreactive neurons and fibers for a hypothalamic neuropeptide precursor related to secretin. *Society for Neuroscience Abstracts* 23: 2032.
- Peyron, C., S.W. Wurts, H. K. Srere, H.C. Heller, D.M. Edgar, and T. S. Kilduff (1998). mRNA levels of Brain-Derived Neurotrophic Factor increase in the cerebral cortex after sleep deprivation. *Sleep Research* 21:9.
- Peyron, C., S. W. Wurts, H. K. Srere, H. C. Heller, D. M. Edgar and T. S. Kilduff. mRNA level of Brain Derived-Neurotrophic Factor (BDNF) increases in several brain regions after sleep deprivation (1998). *Society for Neuroscience Abstracts* 24:1430.
- (7) SCIENTIFIC PERSONNEL: Thomas S. Kilduff, Ph.D. Christelle Peyron, Ph.D.
- (8) Report of INVENTIONS:
- L. de Lecea, T.S. Kilduff, P.E. Foye, P.E. Danielson, V.T. Gautvik, F.E. Bloom, K.M. Gautvik, and J.G. Sutcliffe. Hypothalamus-specific polypeptides. Provisional application submitted 8/2/96. International patent application filed 8/1/97. Publication of the international application with the international search report, 2/12/98.

(9) BIBLIOGRAPHY

- Bitting L, Watson FL, O'Hara BF, Kilduff TS, Heller HC (1999) HSP70 expression is increased during the day in a diurnal animal, the golden-mantled ground squirrel Spermophilus lateralis. Mol Cell Biochem 199:25-34.
- de Lecea L, Kilduff TS, Peyron C, Gao X-B, Foye PE, Danielson PE, Fukuhara C, Battenberg ELF, Gautvik VT, Bartlett II FS, Frankel WN, van den Pol AN, Bloom FE, Gautvik KM, Sutcliffe JG (1998) The hypocretins: hypothalamus-specific peptides with neuroexcitatory activity. Proc Natl Acad Sci U S A 95:322-327.
- Hagan JJ, Leslie RA, Patel S, Evans ML, Wattam TA, Holmes S, Benham CD, Taylor SG, Routledge C, Hemmati P, Munton RP, Ashmeade TE, Shah AS, Hatcher JP, Hatcher PD, Jones DN, Smith MI, Piper DC, Hunter AJ, Porter RA, Upton N (1999) Orexin A activates locus coeruleus cell firing and increases arousal in the rat. Proc Natl Acad Sci U S A 96:10911-10916.
- Horvath TL, Peyron C, Diano S, Ivanov A, Aston-Jones G, Kilduff TS, van Den Pol AN (1999) Hypocretin (orexin) activation and synaptic innervation of the locus coeruleus noradrenergic system. J Comp Neurol 415:145-159.
- Kilduff TS, de Lecea L, Usui Ĥ, Sutcliffe JG (1998) Isolation and identification of specific transcripts by subtractive hybridization. In: *Molecular Regulation of Conscious States* (Lydic R, ed), pp 103-118. Boca Raton, FL: CRC Press.
- Kilduff TS, Peyron C (2000) The hypocretin/orexin ligand-receptor system: Implications for sleep and sleep disorders. Trends Neurosci 23:359-365.
- O'Hara BF, Watson FL, Srere HK, Kumar H, Wiler SW, Welch SK, Bitting L, Heller HC, Kilduff TS (1999) Gene expression in the brain across the hibernation cycle. J Neurosci 19:3781-3790.
- Peyron C, Faraco J, Rogers W, Ripley B, Overeem S, Charnay Y, Nevsimalova S, Aldrich M, Reynolds D, Albin R, Li R, Hungs M, Pedrazzoli M, Padigaru M, Kucherlapati M, Fan J, Maki R, Lammers GJ, Bouras C, Kucherlapati R, Nishino S, Mignot E (2000) A mutation in a case of early onset narcolepsy and a generalized absence of hypocretin peptides in human narcoleptic brains. Nat Med 6:991-997.
- Peyron C, Tighe DK, van den Pol AN, de Lecea L, Heller HC, Sutcliffe JG, Kilduff TS (1998a) Neurons containing hypocretin (orexin) project to multiple neuronal systems. J Neurosci 18:9996-10015.

- Peyron C, Wurts SW, Srere, H. K., Heller HC, Edgar DM, Kilduff TS (1998b) mRNA levels of Brain-Derived Neurotrophic Factor increase in the cerebral cortex after sleep deprivation. Sleep Research 27:9.
- Piper DC, Upton N, Smith MI, Hunter AJ (2000) The novel brain neuropeptide, orexin-A, modulates the sleep-wake cycle of rats. Eur J Neurosci 12:726-730.
- Sakurai T, Amemiya A, Ishii M, Matsuzaki I, Chemelli RM, Tanaka H, Williams SC, Richardson JA, Kozlowski GP, Wilson S, Arch JR, Buckingham RE, Haynes AC, Carr SA, Annan RS, McNulty DE, Liu WS, Terrett JA, Elshourbagy NA, Bergsma DJ, Yanagisawa M (1998) Orexins and orexin receptors: a family of hypothalamic neuropeptides and G protein-coupled receptors that regulate feeding behavior. Cell 92:573-585.
- Thannickal T, Moore R, Y., Nienhuis R, Ramanathan L, Gulyani S, Aldrich M, Cornford M, Siegel JM (2000) Reduced number of hypocretin neurons in human narcolepsy. Neuron 27:469-474.
- van den Pol AN, Gao XB, Obrietan K, Kilduff TS, Belousov AB (1998) Presynaptic and postsynaptic actions and modulation of neuroendocrine neurons by a new hypothalamic peptide, hypocretin/orexin. J Neurosci 18:7962-7971.